

Theoretical Biophysics A Computational Approach Concepts, Models, Methods and Algorithms Static and Temporal Networks

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Introduction

Introduction I









Neural network Source:https://en.wikipedia.org/wiki/Biological_neural_network network

Cytoskeleton

Source: Wikipedia

Source: Wikipedia

https://en.wikipedia.org/wiki/Gene co-

expression network

https://de.wikipedia.org/wiki/Cytoskelett

Introduction II





Figure 2 Network members (n = 89) viewed by their connection through a bar associated with gonorrhoea acquisition. A prefix to the unique identifier of "" designates a male and "f" indicates a female sexual partner. Bar patrons possessed significantly higher information centrality measures compared to non-patrons (dable 3).

Sexual network analysis of a gonorrhoea outbreak

Image taken from: P De, A E Singh, T Wong, et al. doi: 10.1136/sti.2003.007187 2004 80: 280-285 Sex Transm Infect

Apotosis





Image taken from Bentele M, Lavrik I, Ulrich M, Stosser S, Heermann DW, Kalthoff H, Krammer PH, Eils R (2004). Mathematical modeling reveals threshold mechanism in CD95-induced apoptosis. J Cell Biol 166, 839-851

Gene Interaction Network







Graph Models





Figure 1: Square lattice.

Graph Models: Lattice Examples II





Figure 2: Petersen graph is a graph where no pair of vertices are separated by more than two edges.

Graph Models: Lattice Examples III





Cayley tree with degree 3



Hexagonal graph with degree 3

Figure 3: Cayley tree and hexagonal lattice.

Graph Models I



A graph consists of a nonempty set of **points** or **vertices**, and a set of **edges** that link together the vertices. A graph may be of two many forms: **directed** or **undirected**. In a directed graph the direction of any given edge is defined.

An edge in a graph that joins two vertices is said to be **incident** to both vertices. The **degree** of a vertex is determined by the number of distinct edges that are incident to the vertex.

Two edges in a graph are **adjacent** if they connect to the same vertex.

A loop is an edge that links a vertex to itself.

A **path** through a graph is a traversal of consecutive vertices along a sequence of edges. A **cycle** is a path in which the initial vertex of the path is also the terminal vertex of the path.

- A graph $G_1 = (V_1, E_1)$ is a subgraph of G = (V, E) if $V_1 \subset V$ and $E \subset E_1$.
- In general each edge may be associated with a direction and weight $w_i \in \mathbf{R}$.
- In a **directed graph** we attach a direction to each edge e_s^d . $e_s^d = (v_i, v_j)$ means that the edge e_s starts at node v_i and ends at node v_j .

Graph Models II



In an undirected graph the order in which nodes are written does not matter

$$e_s^n = (v_i, v_j) = (v_j, v_i)$$
 (1)

- We allow for $v_i = v_j$. Such an edge is said to be a **one-edged loop** attached to v_i .
- It is possible to allow for more than one edge between nodes v_i and v_j .
- If a graph contains neither multiple edges between pairs of nodes nor loops, then the graph is called simple.
- For a simple graph the number of edges is at most

$$M^{\max} = \frac{N(N-1)}{2} . \tag{2}$$

In this case the graph is called **fully connected**.

■ The degree *d_i* is number of edges incident to a node *v_i*. In a directed graph we distingiush between the **in-degree** *d_i*^{IN} and the **out-degree** *d_i*^{OUT}, ie., the number of nodes ending on or starting from node *v_i*.

Graph Models III



$$\Gamma(v_i) := \{v_j | v_j \in V \land (v_i, v_j) \in E\}$$
(3)

clearly the degree (in-degree) is also the size of the neighbourhood

$$d_i = |\Gamma(v_i)| . \tag{4}$$

In all graphs we have

$$\sum_{i} d_{i} = 2M, \quad M = |E|.$$
(5)

For directed graphs $\sum_i d_i = M$

• The average degree \overline{d} of a graph is defined as

$$\bar{d} := \frac{1}{N} \sum_{i=1}^{N} d_i$$
 (6)

For directed graphs we have

$$\frac{1}{N}\sum_{i=1}^{N}d_{i}^{|\mathsf{N}|} = \frac{1}{N}\sum_{i=1}^{N}d_{i}^{\mathsf{OUT}}.$$
(7)



Graph Models IV



■ The degree distribution is defined by

$$P(k) := \frac{1}{N} \sum_{i=1}^{N} \delta_{d_i,k} \quad \text{for} k = 0, 1, 2, \dots$$
(8)

The degree distribution summarizes information about the local environment of a graph

- $P^{IN(k)}$ can be different from $P^{OUT(k)}$.
- A set of *k* nodes *C* = {*v*₁,...,*v_k*} where each node in *C* can be reached from other nodes in *C* but not from any node outside of *C* is called a **connected component of size** *k*.
- For a simple graph the number of connected components K is given by

$$K \ge N - M \tag{9}$$

- If there is more than one path between a pair of nodes $v_i, v_j \in V$, then the graph contains closed paths or loops.
- In an undirected simple graph, if there is precisely one path between each pair of nodes v_i, v_i ∈ V, then there cannot be any loop and the graph is called a tree.

Graph Models V



- If a graph consist of several components, each of which is a tree, the graph os called a **forrest**.
- A spanning tree T of a connected graph with nodes $V_T = V_G$ and edges $E_T \subseteq E_G$ is a graph such that (V_T, E_T) is a tree.

Distances and Diameter I



 $I_{ij} := \min\{x_{ij} | x_{ij} \text{ is the length of a path from } v_i \text{ to } v_j \text{ along } e_s \in E\}$ (10)

- \blacksquare If there is no path, then we set $\mathit{I_{ij}} = \infty$
- $\blacksquare \text{ In general } I_{ij} \neq I_{ji}$
- The diameter of a graph is defined as the maximum distance between two nodes in the graph

$$D = \max\{l_{ij} | v_i, v_j \in V\}$$
(11)

For a disconnected graph we have $D = \infty$. This can be made finite by defining D to be the diameter of the largest connected component.



Distances and Diameter II

■ The clustering coefficient measures the probability that two nodes v_j and v_k , which are both neighbours of v - i ((v_i, v_j) , $(v_i, v_k \in E)$ are themselves connect by an edge $(v_i, v_k) \in E$

$$c_i := \frac{2\eta_i}{d_i(d_i - 1)} \quad \text{for} d_i \ge 2 \tag{12}$$

where η_i is the number of edges among the nodes connected to v_i .

The average path length is defined as

$$\bar{l} := \frac{2}{N(N-1)} \sum_{i=1}^{N} \sum_{j=1}^{N} l_{ij}$$
(13)

 $I_{ii} = 0$ by definition.

■ The distribution of graph distances is defined as

$$\lambda(l) := \frac{2}{N(N-1)} \sum_{i=1}^{N} \sum_{j=1}^{N} \delta_{l_{ij},l} \quad l = 1, 2, \dots$$
(14)







Figure 4: Example: Erdös-Renyi Model, A p = 0.1, B p = 0.3, C p = 1.

- Graphical display of dependence structure between multiple interacting quantities (expression levels of different genes).
- Probabilistic semantics: Fits the stochastic nature of both the biological processes and noisy experiments. Capable of handling noise and estimating the confidence in the different features of the network.

Random Graphs: Erdös-Renyi-Model II



- Due to lack of data: Extract features that are pronounced in the data rather than a single model that explains the data.
- Random variable X_i = measured expression level of gene *i* represented by nodes.
- Edges = regulatory interactions between genes.
- Define the functional form of the conditional distributions (e.g. multinomial for discrete variables, linear Gaussian for continuous variables).
 - Find the best network structure *S*
 - Given a network structure, find the best set of parameters for the conditional distributions (the most probable structure/parameter vector given the data)

Random Graphs: Erdös-Renyi-Model III





Figure 5: Fraction of largest cluster for the Erdös-Renyi model.

Erdös-Rényi Model

Let G be a graph with N nodes. Any two nodes have an edge with probability p. Each of the (N-1)N/2 edges appears independently with probability p.

Random Graphs: Erdös-Renyi-Model IV

• The total number of edges is a random variable with expectation value

$$< n >= p[N(N-1)/2]$$
. (15)

If G_0 is a graph with nodes $V_1, V_2, ..., V_N$ and *n* edges the probability of realizing this graph is

$$P(G_0) = p^n (1-p)^{N(N-1)/2-n} .$$
(16)

A graph $G_1 = (V_1, E_1)$ is a **subgraph** of G = (V, E) if $V_1 \subseteq V$ and $E_1 \subseteq E$.

Examples

- cycles are closed loops of k edges such that every two consecutive edges and only those have a common node.
- **complete subgraphs** (of order k) contains k nodes and all of the possible k(k-1)/2 edges.



Random Graphs: Erdös-Renyi-Model V



Question: Is there a critical probability that marks the appearance of arbitrary subgraphs consisting of k nodes l edges?

In a random graph with connection probability p the degree k_i of node i follows a binomial distribution with parameters N-1 and p

$$P(k_i = k) = C_{N-1}^k p^k (1-p)^{N-1-k} .$$
(17)

There are C_{N-1}^k equivalent ways of selecting the k end points.

• For $i \neq j$ two different nodes, $P(k_i = k)$ and $P(k_j = k)$ are close to being to independent random variables.

Random Graphs: Erdös-Renyi-Model VI

Let X_k be the number of nodes with degree k. Our goal is to determine the probability that X_k takes on a given value

$$P(X_k = r) \tag{18}$$

$$\langle X_k \rangle = NP(k_i = k) = \lambda_k = NC_{N-1}^k p^k (1-p)^{N-1-k}$$
 (19)

Assume $P(X_k = r)$ approaches a Poisson distribution

$$P(X_k = r) = e^{\lambda_k} \frac{\lambda_k^r}{r!} .$$
⁽²⁰⁾

Thus the degree distribution of a random graph is a binomial distribution

$$P(k) = C_{N-1}^{k} p^{k} (1-p)^{N-1-k} .$$
⁽²¹⁾

For large N this can be replaced by a Poisson distribution

$$P(k) = e - pN \frac{(pN)^k}{k!} = e^{\langle k \rangle} \frac{\langle k \rangle^k}{k!} .$$
(22)



Let G = (V, E) be a directed acyclic graph. We assume that the vertices $i \in V$ $(1 \le i \le n)$ represent for example genes and correspond to random variables x_i . For each y_i we define a conditioned probability

$$P(x_i | \mathsf{parent}(x_i)) = P(x_i | P_a(x_i))$$
(23)

and the joint probability distribution

$$P(x_1,...,x_n) = \prod_{i=1}^n P(x_i | P_a(x_i)) .$$
(24)

We define a parameter θ to be the set of conditioned probabilities.

From the graph shown in Figure 6 representing the Bayesian network we can read off the joint probability distribution

$$P(x_1, ..., x_n) = P(x_1)P(x_2)P(x_3|x_1, x_2)P(x_4|x_2)P(x_5|x_3, x_4)$$
(25)

and θ is the set of local conditional probabilities.



Bayesian Networks II





Figure 6: A simple Bayesian network.

Bayesian Networks III



Let x be distributed as $x \sim f_{\theta}(x)$ with parameter θ . Our aim is to estimate the parameter θ , given that there are iid observations $\{x_i\}$ of the random variable x.

Example:

Let $\{x_i\}$ be a sequence of results of rolling a dice such that $x_i = 1$ denotes head and $x_i = 0$ tail. The dice is unfair so that we have for the probability that the random variable takes on the result $x_i = 1$ is $P(x_i = 1) \neq 1/2$. Define the parameter $\theta = P(x_i = 1)$. We need an estimate for θ . Of course, we know that we are dealing with a binomial distribution and

$$P(x=1) = p, \quad P(x=0) = q, \quad q = 1 - p.$$
 (26)

How many successful realizations k do we have after we have tried *n*-times? This is given by the binomial distribution (recall that the reverse question gives the negative binomial distribution).

Without any prior information we can use the maximum likelihood method to estimate the parameter θ . Let $x_1, ..., x_n$ denote the observations, with H + T = n where H is the number of trails we have obtained head, and T the number of trails we have obtained tail. Then

Bayesian Networks IV



$$\hat{\theta} = \operatorname{argmax} f_{\theta}(y_i) \quad i = 1, \dots n$$
 (27)

$$= \operatorname{argmax} \theta^{H} (1 - \theta)^{T}$$
(28)

$$= \frac{H}{H+T}$$
(29)

which corresponds to the empirical frequency of the sample for the event H. Even if we have prior information in the sense that we have a previous set of experiments where the number of heads was 5 and the number of tails was 10, we would obtain

$$\hat{\theta} = \frac{5+H}{5+H+10+T}$$
(30)

which is irrelevant if the number of total experiments n = T + H goes to infinity.

But now suppose we are not sure that we measured 5 heads and 10 tails and can only tell that presumably the probability with which we obtained head was 1/4. For this uncertainty we assume a prior distribution for *theta* $P(\theta)$

$$P(\theta) = \beta(5, 10) = \frac{\Gamma(5)\Gamma(10)}{\Gamma(5+10)}$$
(31)

Bayesian Networks V



where we have assumed that the prior is distributed as a beta distribution

$$\beta(p,q) \sim \frac{\Gamma(5)\Gamma(10)}{\Gamma(5+10)} = \int_0^1 x^{p-1} (1-x)^{q-1} dx .$$
(32)

Assume that we make new observations. We use the Bayesian law to compute the posterior distribution

$$P(\theta|x) = \frac{P(x|\theta)P(\theta)}{P(x)}$$
(33)

$$= \frac{P(x|\theta)P(\theta)}{\int P(x|\theta)P(\theta)d\theta}$$
(34)

where $P(x|\theta)$ is the likelihood function. In the new experiment we find 50 heads and 50 tails so that we obtain for the likelihood function

$$P(x|\theta) = \theta^{50} (1-\theta)^{50}$$
(35)

from which we obtain the posterior distribution

Bayesian Networks VI



$$P(\theta|x) = \frac{\theta^{59}(1-\theta)^{69}}{\int \theta^{59}(1-\theta)^{69}d\theta} .$$
(36)

Note that the posterior function is the same as the prior function. In such a case we call the distribution conjugate. Thus the beta distribution is the conjugate to the binomial distribution.

The result of a Bayesian analysis is a posterior distribution and not a single value! This distribution can be used to make predictions.



Graphic representation of a joint distribution over a set of random variables A, B, C, D, E.

P(A, B, C, D, E) = P(A) * P(B) * P(C|A) * P(D|A, B) * P(E|D)



Figure 7: Nodes represent gene expression while edges encode the interactions (cf. inhibition, activation).

Bayesian Networks for Gene Expression Data II



- Given a set of random variables $X = (X_1, ..., X_n)$, a Bayesian network is defined as a pair $BN = (S, \theta)$, where
 - S is a directed acyclic graph (DAG), which is a graphical representation of the conditional independencies between variables in X
 - $\blacksquare \ \theta$ is the set of parameters for the conditional probability distributions of these variables.
 - In a Bayesian network, the probability of a state $x = (x_1, x_2, ..., x_n)$ is factored as

 $P(x) = P(x_1 | pa(x_1)) P(x_2 | pa(x_2)) \dots P(x_n | pa(x_n)),$

where pa(x) denotes the parents of node x in the graph S

- A Bayesian network should be a DAG (Direct Acyclic Graph).
- Random variable X_i = measured expression level of gene i. Arcs = regulatory interactions between genes.
- However, there are lots regulatory networks having directed cycles.
- Solve this by expanding into the time direction.

Bayesian Networks for Gene Expression Data III





Figure 8: Use DBN (Dynamic Bayesian Networks: BN with constraints on parents and children nodes) for sequential gene expression data.

Bayesian Networks for Gene Expression Data IV



We are looking for a Bayesian network that is most probable given the data D (gene expression)

$$BN^* = \operatorname{argmax}_{BN} \{ P(BN|D) \}$$
(37)

where

$$P(BN|D) = \frac{P(D|BN)P(BN)}{P(D)} .$$
(38)

There are many networks. An exhaustive search and scoring approach for the different models will not work in practice (the number of networks increases super-exponentially, $O(2^{n^2})$ for dynamic Bayesian networks)

 $\ensuremath{\mathsf{Idea:}}$ Sample the networks such that we eventually have sampled the most probable networks

Monte Carlo

Recall detailed balance condition for Monte Carlo

$$P(BN_{old}|D)P(BN_{old} \rightarrow BN_{new}|D) = P(BN_{new}|D)P(BN_{new} \rightarrow BN_{old}|D) .$$

Let us look at

$$P(BN|D) = \frac{P(D|BN)P(BN)}{P(D)}$$

■ Assume *P*(*BN*) is uniformly distributed (*We could incorporate knowledge*).

Bayesian Networks for Gene Expression Data V



- Choose next BN with probability P(BN_{new}).
- Accept the new BN with the following Metropolis-Hastings accept/rejection criterion:

$$\begin{split} P &= \min\left\{1, \frac{P(BN_{\mathsf{new}}|D)P(BN_{new} \rightarrow BN_{old}|D)}{P(BN_{\mathsf{old}}|D)P(BN_{old} \rightarrow BN_{new}|D)}\right\} \\ &= \min\left\{1, \frac{P(D|BN_{\mathsf{new}})P(BN_{\mathsf{new}})P(D)}{P(D|BN_{\mathsf{old}})P(BN_{\mathsf{old}})P(D)}\right\} \\ &= \min\left\{1, \frac{P(D|BN_{\mathsf{new}})P(BN_{\mathsf{new}})}{P(D|BN_{\mathsf{old}})P(BN_{\mathsf{old}})}\right\} \\ &= \min\left\{1, \frac{P(D|BN_{\mathsf{new}})P(BN_{\mathsf{new}})}{P(D|BN_{\mathsf{old}})P(BN_{\mathsf{old}})}\right\} \\ &= \min\left\{1, \frac{P(D|BN_{\mathsf{new}})}{P(D|BN_{\mathsf{old}})}\right\} . \end{split}$$

Discrete model

- Even though the amount of mRNA or protein levels, for example, can vary in a scale that is most conveniently modeled as continuous, we can still model the system by assuming that it operates with functionally discrete states
 - activated / not activated (2 states)
 - under expressed / normal / over expressed (3 states).
- Discretization of data values can be used to compromise between the
 - averaging out of noise,

Bayesian Networks for Gene Expression Data VI



- accuracy of the model,
- complexity/accuracy of the model/parameter learning.
- Qualitative models can be learned even when the quality of the data is not sufficient for more accurate model classes.
- Let *N_{ijk}* be the number of times we observe variable/node *i* in state *k* given parent node configuration *j*.
- Summarize the number of total number of observations for variable *i* with parent node configuration *j*,

$$N_{ij} = \sum_{k=1}^{r_j} N_{ijk}$$

- Since our states are discrete we use a multinomial distribution
- the ML estimate of multinomial probabilities is obtained by the normalized counts

$$\hat{\theta}_{ijk} = \frac{N_{ijk}}{N_{ij}}$$

A convenient prior distribution to choose for the parameters θ is given by the Dirichlet distribution

$$(\theta_{ij1}, ..., \theta_{ijr_i}) \sim \mathsf{Dirichlet}(\alpha_{ij1}, ..., \alpha_{ijr_i})$$
.
Bayesian Networks for Gene Expression Data VII



The Dirichlet distribution has PDF

$$f(\theta_{ij1},...\theta_{ijr_i};\alpha_{ij1},...\alpha_{ijr_i}) = \frac{1}{B(\alpha_{ij})} \prod_{i=1}^{r_i} \theta_{ijr_i}^{\alpha_{ijr_i}-1}$$

with $\theta_{ijr_i} \ge 0, \sum_i \theta_{ijr_i} = 1$ and hyperparameters $\alpha_{ijr_i} \ge 0, \alpha_{ij} = \sum_k \alpha_{ijr_i}$.

The normalization constant, the Beta function, can be expressed using the gamma function

$$B(\alpha_{ij}) = \frac{\prod_{k=1}^{r_i} \Gamma(\alpha_{ijr_i})}{\Gamma(\alpha_{ij})}$$

- The convenience arises from the fact that the distribution is conjugate to the multinomial distribution, i.e., if $P(\theta)$ is Dirichlet and $P(X|\theta)$ is multinomial, then $P(\theta|X)$ is Dirichlet as well.
- The multinomial distribution is given (for $N_{ij} = \sum_k N_{ijk}$) by

$$f(N_{ij1}, \ldots, N_{ijr_i} | N_{ij}, \theta_{ij1}, \ldots, \theta_{ijr_i}) = \frac{N_{ij1}}{N_{ij1}! \cdots N_{ijr_i}!} \theta_{ij1}^{N_{ij1}} \cdots \theta_{ijr_i}^{N_{ijr_i}} \ .$$

and is the distribution of observations in r_i classes if N_{ij} observations are selected as outcomes of independent selection from the classes with probabilities θ_{ijk} , $k = 1, ..., r_i$.

Structural Properties

Bayesian Networks for Gene Expression Data VIII



- In order to get reliable results we can focus on features that can be inferred
 - for example, we can define a feature, an indicator variable f with value 1 if and only if the structure of the model contains a path between nodes A and B.
 - Looking at a set of models S with a good fit we can approximate the posterior probability of feature f by

$$P(f|D) = \sum_{S} f(S)P(S|D) .$$

- With gene regulatory networks, one can look for only the most significant edges based on the scoring.
- A Markov chain is defined over Bayesian nets so that it approaches a steady-state distribution as it is being run, and the probabilities of the states (networks) correspond to their posterior probability.
- Individual nets are created as states in the chain and after (assumed) convergence, samples *S_i* are taken
- Posterior probability of an edge can then be approximated with

$$P(f(S)|D) \approx \frac{1}{n} \sum_{i=1}^{n} f(S_i)$$

Bayesian Networks for Gene Expression Data IX



• To work out the Monte Carlo Method to generate networks we first have to compute P(D|S)

$$P(D|S) = \int_{\theta} P(D|\theta, S)P(\theta|S)d\theta$$

= ...
$$= \prod_{i=1}^{n} \prod_{j=1}^{q_i} \frac{\Gamma(\alpha_{ij})}{\Gamma(\alpha_{ij} + N_{ij})} \prod_{k=1}^{r_i} \frac{\Gamma(\alpha_{ijk} + N_{ijk})}{\Gamma(\alpha_{ijk})}.$$

■ Monte Carlo moves: ADD, REMOVE, REVERSE edge in network.

Weight Matrix Model I



A weight matrix model considers the interactions between all combinations of genes. A weight matrix [1] consists of $n \times n$ weight values, each of which indicates the influence of one specific gene on another. A positive value for $w_{i,j}$ models gene j stimulating the expression of gene i. A negative value models repression, while a value of zero indicates that gene j does not influence the transcription of gene i.

The expression state of a transcriptional regulatory network containing *n* genes (with discrete states) is represented by a vector $\mathbf{u}(t)$. The net regulatory effect, of gene *j* on gene *i* is the expression level of *j*, i.e., $u_j(t)$, times its regulatory influence on *i* which is determined by the weight $w_{i,j}$. The total regulatory input to *i*, i.e. $r_i(t)$, is derived by summing over all the genes in the system

$$r(i) = \sum_{j} w_{ij} u_j(t) .$$
(39)

The response of each gene to the regulatory input in this model is calculated with a dose-response function

$$x_i(t+1) = \frac{1}{1 + e^{-(\alpha_i r_i(t) + \beta_i)}}$$
(40)

Weight Matrix Model II

where α_i and β_i are two gene specific constants that define the shape of the dose-response curve for gene *i*. This assumes that each gene has a static dose-dependent response to activating and repressing regulatory influences.

The constants can be incorporated into the weight matrix, replacing W with Z

$$Z_{ij} = \alpha_i w_{ij} . \tag{41}$$

We further define a column weight $z_{i0} = \beta_i$ and $u_0(t) = 1$. Thus the vector of net regulations $(\mathbf{r}(t))$ now is $\mathbf{s}(t)$

$$s_i(t) = \sum_j u_j(t) z_{ij} \tag{42}$$

and

$$x_i(t+1) = \frac{1}{1 + e^{-s_i(t)}}$$
(43)



(11)



$$u_i(t+1) = m_i x(t) = \frac{m_i}{1 + e^{-\sum_j z_{ij} u_j(t)}}$$
(44)

here m_i is the maximal expression level for gene *i* used to get the real expression output for *i*, i.e. $u_i(t + 1)$.



A Boolean network is one of the simplest model for the behaviour of genomic network. Such network consists of n nodes (e.g. representing genes) which can either be repressed or expressed (the node has state 0 or 1, respectively). The dynamics of the network is determined by a list of n (Boolean) functions which each receive input from k specified nodes. Every node has its own specific function, which can determine its next state from the current states of all the input nodes.

A **Boolean network** [2, 3] G(V, F) is thus defined by a set of nodes $V = \{x_1, ..., x_N\}$ and a list of Boolean functions $F = (f_1, ..., f_k)$ corresponding to the edges. In these models, gene expression is quantized to just two levels: ON and OFF (1 or 0). The state of a node (gene) is completely determined by the values of other nodes at time *t* by means of underlying logical Boolean functions. The model is represented in the form of directed graphs. Each x_i represents the state (expression) of gene *i*, where $x_i = 1$ represents the fact that gene *i* is expressed and $x_i = 0$ means it is not expressed.

The time evolution of the network is described by

$$x(t+1) = f_i(x_{j_1}(t), ..., x_{j_{k_i}}(t))$$
(45)

where k_i denotes the connectivity of the node *i*. For the average connectivity

Random Boolean Networks II

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we have

$$\langle K \rangle >= \frac{1}{N} \sum_{i=1}^{N} k_i .$$
(46)

The list of Boolean functions F represents the rules of regulatory interactions between genes. The tuple (G, F) is one realization. In the *quenched model* one realizations is chosen at kept fixed for all times. In the *annealed model* a new realization is chosen at random at very time step.

Any given gene transforms its inputs (regulatory factors that bind to it) into an output, which is the state or expression of the gene itself. All genes are assumed to update synchronously or asynchronously in accordance with the functions assigned to them and this process is then repeated. Despite the simplicity of the model, they can capture a number of essential features of real genomic networks.

Random Boolean Networks III





Figure 9: A random network with 5 nodes and 10 edges and one with 16 nodes and 28 edges.



Asynchronous random boolean networks (ARBN) incorporate all the cases in which at each time point a single node is selected in order to be updated. The node to be updated can be chosen at random or according to a deterministic rule based:

- clock scheme [4, 5],
- cyclic scheme [6],
- random independent scheme [7], and
- random order scheme [7].

Consider the example shown in Figure 10. The assignment of values to nodes made in the truth table fully describes the state of the model at any given time. The change of model state over time is fully defined by the functions in F. If we assign initial values to the nodes, all further states are determined. Consider the time evolution of the states as a trajectory. Since the number of possible states is finite, all trajectories eventually end up in single *steady states*, or a *cycle of steady states*. In this context we define an *attractor* of a trajectory as a single steady state, or a cycle at the end of the trajectory. The *basin of attraction* for a specific attractor is a set of all trajectories leading to it. States in gene networks are often characterized by stability in the sense that small changes in value of a few nodes do not change the attractor.

Random Boolean Networks V





Figure 10: A sample boolean network.

At each time point t the system can be in one of the 2^N possible states. We have 2^{2^k} possible logic functions per node which yield

possible nets =
$$N_F = \left(\frac{2^{2^k}N!}{(N-k)!}\right)^N$$
. (47)

The geometry of the network strongly influences the behaviour. We can distinguish between $% \left({{{\left[{{{\rm{s}}_{\rm{c}}} \right]}}} \right)$

Random Boolean Networks VI



- irregular networks,
- regular networks like lattices where for example we have |x_{j1}(t),...,x_{jk_i(t)}| = 2d neighbours on a simple hypercubic lattice.

From the statistical mechanics point of view we have to specify an ensemble of the coupling functions, i.e. each must be assigned a weight. For this we can consider the following options

- equal weight ensemble
- magnetization biased
- forcing functions
- additive functions

Random Boolean Networks VII



In the equal weight ensemble we assign every coupling function the same weight $1/N_F$.

In the magnetization bias situation the probability of occurrence of a coupling function is p if the result is 0 and 1 - p if the result is 1.

In the forcing function ensemble the value of the function is determined if one of the arguments $m \in \{1, ..., k\}$ takes on a predetermined value. For example $x_m = 0$. The value of the function is not determined if $x_m = 1$.

And for the additive function ensemble we have

$$x_i(t+1) = \Theta(f_i(t)) \tag{48}$$

and

$$f(t) = h + \sum_{j=1}^{N} c_{ij} x_j(t)$$
(49)

where h is a bias.

Now let us look at the response of the system if we change conditions. Let



$$\Sigma_0 = \{x_1(0), ..., x_N(0)\}$$
(50)

denote the starting state and

$$\hat{\Sigma}_{0} = \{x_{1}(0), ..., x_{N}(0)\}$$
(51)

another starting state which differs from Σ_0 only in a few nodes. To describe the difference we introduce the Hamming distance

$$D(t) = \sum_{i=1}^{N} (x_i(t) - \hat{x}(t))^2$$
(52)

to describe the difference in the evolution of the network, given two initial conditions. Clearly, if the network is such that any local discrepancy remains localized, then we do expect the Hamming distance to remain finite in the thermodynamic limit. If, on the other hand the discrepancy (or damage) can be propagated to almost every vertex, then we expect the Hamming distance to diverge.

Random Boolean Networks IX

Assume that we are dealing with the uniform distribution for the coupling functions. On average, a change of a single vertex will affect the argument of k functions. Hence kD(0) functions are affected. Each of these is affected with a probability 1/2 so that D(1) = kD(0) and in general

$$D(t) = \left(\frac{k}{2}\right)^t D(0) .$$
(53)

We can thus distinguish three phases

- chaotic phase for $k_c > 2$
- frozen for $k_c < 2$
- critical for $k_c = 2$





Random Boolean Networks X



In the last case the state of the system will be dominated by fluctuations. Let us now look at the case where

$$f_i = \begin{cases} 0 & \text{with probability } p \\ 1 & \text{with probability } 1 - p \end{cases}$$
(54)

For a given p and a given connectivity k we will have critical values

$$k_c(p)$$
 and $p_c(k)$. (55)

Let

$$a(t) \equiv 1 - D(t)/N \tag{56}$$

be the probability that two vertices have the same value in Σ_t and $\hat{\Sigma}$. The probability that the arguments of the functions f_i have the same value is given by

$$\rho_k = [a(t)]^k \tag{57}$$

Random Boolean Networks XI



The overlap is the same in the next time step if the arguments of the coupling functions are identical. This occurs with probability ρ_k . The overlap is also the same if the arguments of the coupling functions are also different, which occurs with probability $1 - \rho_k$ but the values are nevertheless the same, which occurs with probability 2p(1 - p). Altogether we have

$$a(t+1) = 1 - (1 - \rho_k) 2p(1-p)$$
(58)

$$= 1 - \frac{1 - [a(t)]^k}{k_c}$$
(59)

where we have used

$$k_{c} = \frac{1}{2p(1-p)}$$
(60)
$$p_{c} = \frac{1}{2} \pm \sqrt{\frac{1}{4} - \frac{1}{k}} .$$
 (61)

Clearly the recursion relation 59 has a trivial fixpoint given by



$$a^* = 1$$
. (62)

What kind of fixpoint is this? To answer this question we perturb the fixpoint slightly with δa_t . Then coming from below we have

$$1 - \delta a_{t+1} = 1 - \frac{1 - [1 - \delta a_t]^k}{k_c}$$
(63)

yielding

$$\delta a_{t+1} \approx \frac{k}{k_c} \delta a_t . \tag{64}$$

The trivial fixpoint is unstable if $k/k_c > 1$. The resulting phase diagram can be seen in Figure 11.

Random Boolean Networks XIII





Figure 11: The phase diagram is only shown for $0 \le p \le 0.5$.



So far we could attach a definite scale to the network. We shall now investigate networks that are scale free. For this we use the following connectivity distribution

$$P(k) = \frac{1}{\zeta(\gamma)} k^{-\gamma} \quad , \quad \gamma > 1 \quad .$$
 (65)

with

$$\zeta(\gamma) = \sum_{k=1}^{\infty} k^{-\gamma} \tag{66}$$

being the Riemann Zeta-function.

The condition that P(k) is normed requires that $\gamma > 1$. For the first moment we obtain

$$\langle k \rangle = \sum_{k=1}^{\infty} k P(k) = \begin{cases} \infty & \text{if } 1 < \gamma \le 2\\ \zeta(\gamma - 1)/\zeta(\gamma) & \text{if } \gamma > 2 \end{cases}$$
(67)

Scale-Free Networks II

As done in the previous section we considered the case of the annealed model. Every element receives k inputs with probability P(k)

$$a(t+1) = G(a(t)) \tag{68}$$

and recall that a(t) = 1 - D(t)/N. The average probability that k = 1, 2, ... controlling elements of the coupling function f are identical is

$$\mu(a) = \sum_{k=1}^{\infty} a^k P(k) .$$
(69)

From this we can calculate the recursion function G

$$G(a) = 1 - 2p(1-p)[1 - \sum_{k=1}^{\infty} a^k P(k)].$$
(70)

For the fixpoint a^* of Equation 68 we can again analyse the stability by considering $a^* + \delta a^*$. a^* is unstable iff





$$1 = \lim_{a \uparrow 1} \frac{dG(a)}{da}$$
$$= 2p(1-p) \sum_{k=1}^{\infty} kP(k)$$
$$= 2p(1-p)\langle k \rangle.$$
(71)

The phase transition is along this line because the fixpoint is stable for $\lim_{a\uparrow 1}\frac{dG(a)}{da}<1$ and unstable for $\lim_{a\uparrow 1}\frac{dG(a)}{da}>1.$

We shall now analyse the trajectory through phase space in terms of *limit cycles* and *attractors*. For this we switch to the quenched RBN. In this model the coupling functions are independent of time. For every initial condition we can follow Σ_t which surely will reach a state that was visited before. In this case it will cycle. Thus the phase space $\Omega = 2^N$ can be partitioned into cycles and attractors. An attractor can be defined as set of points from the phase space $A_t \equiv {\Sigma_t} \subset \Omega$ which will be mapped onto itself $A_{t+1} = A_t = A_0$. A *basin of attraction* of an attractor A_0 is a subset of Ω with



$$ex \ T < \infty : \Sigma_T \in A_0 \ . \tag{72}$$

Let k = N. Assume that we start with Σ_0 and follow the trajectory. Let q_t denote the probability that the random walk is still not closed after t steps and let p_t denote the probability that the walk terminates (closes) after exactly t steps. If the trajectory is open after t steps then t + 1 different points were visited. Hence there are t + 1 possibilities to terminate the walk in the next step and this happens with the probability $\rho_t = (t + 1)/|\Omega|$). Thus

$$p_{t+1} = \frac{t+1}{|\Omega|} q_t$$
 (73)

The probability that the walk after t + 1 steps is open is

$$q_{t+1} = q_t (1 - \rho_t) = q_t \left(1 - \frac{t+1}{|\Omega|} \right)$$
(74)

with $q_0 = 1$. Due to the fact that the phase space is growing exponentially we can write

Scale-Free Networks V



$$q_t = \prod_{i=1}^t \left(1 - \frac{i}{|\Omega|} \right) \approx \prod_{i=1}^t e^{-i/|\Omega|}$$
(75)
$$= e^{-\sum_i i/|\Omega|} = e^{-t(t+1)/2|\Omega|}.$$
(76)

We can now calculate the average cycle length. Let P(L) denote the probability that a given starting point is in the basin of attraction with cycle length *L*. The closing event happens with equal probability so that

$$P(L) = \sum_{t=L}^{|\Omega|} \frac{p_t}{t}$$
(77)

and for the expectation value

$$\langle L \rangle \approx |\Omega|^{1/2}$$
 (78)





Figure 12: A random network with 5 nodes and 10 edges

Scale-Free Networks VII





Figure 13: Generation of a simple small world random network



Scale free graphs or networks can also arise from growth models. For this we need preferential attachment.

Algorithm 1 Barabasi-Albert-Model

- 1: Start with a disconnected set of m_0 nodes
- 2: New nodes enter the network ate any time step.
- 3: For any new node m' new edges are formed.
- 4: The *m'* new edges connect the nodes with the old nodes. The latter ones are extracted with a probability

$$P(k_i) = \frac{k_i}{\sum_j k_j}$$

Assume that at every time step only one vertex enters. Then we have

Barabasi-Albert-Model II



$$n = m_0 + t \tag{79}$$

$$m' = \frac{1}{2} \sum_{j=1}^{n} k_j = mt$$
 (80)

Assume that verteices enter the graph at a constant rate. Assume too that the degree is a continuous variable. The variation of the degree with time is given by

$$\frac{\partial k_i}{\partial t} = A\Pi(k_i) = A \frac{k_i}{\sum_{j=1}^{m_0+t-1} k_j} = A \frac{k_i}{2mt} .$$
(81)

The constant A is the change in connectivity in one time step, hence A = m. Initially at t_0 , the initial degree is $k(t_0) = m$. It follows that

$$\frac{\partial k_i}{\partial t} = \frac{k_i}{2t} \Rightarrow k_i(t) = m \left(\frac{t}{t_0}\right)^{1/2} . \tag{82}$$

The probability $P(k_i < k)$ that a vertex has a degree k is

Barabasi-Albert-Model III



$$P(k_i < k) = P\left(t_i > \frac{m^2 t}{k^2}\right) .$$
(83)

Since vertices enter at a constant rate, their distribution is uniform in time

$$P(t) = \text{const} \tag{84}$$

Thus

$$\int_{0}^{m_{0}+1} P(t)dt = 1 \Rightarrow P(t) = \frac{1}{m_{0}+t}$$
(85)

and hence

$$P\left(t_{i} > \frac{m^{2}t}{k^{2}}\right) = 1 - P\left(t_{i} \le \frac{m^{2}t}{k^{2}}\right) = 1 - \frac{m^{2}t}{k^{2}(m_{0} + t)}.$$
(86)

It follows that



$$P(k) = \frac{\partial P(k_i > k}{\partial k} = \frac{2m^2t}{m_0 + t} \frac{1}{k^3} .$$
(87)

The distribution floows a power law

$$P(k) \sim k^{-\gamma} \quad , \quad \gamma = 3 \; . \tag{88}$$

Hence the degree-distribution is scale-free.



Gels





Figure 14: Two-dimensional confocal microscopy images of the reconstituted collagen gels analyzed in this study. Image taken from [8].

Gels II



Many systems exhibit a network structure (see Figure 15). The mechanical properties of such networks are often characteristic of a solid even though they being disordered and may even be mostly liquid like. The structure of such a network is described by the following structure parameters. First there is an elastically active network chain between two crosslinks. Then there are dangling chains which are attached to the network by a single point. The gel fraction includes all the material attached to the network.

Let us look at the classical model of gelation proposed by Stockmayer and Flory [9–11] for the sol gel transition. The model assumes monomers having f valences which can bind (simple) to other monomers to form a network. Not all valences are bound. We assume that each valence has probability of $p(T, c_i, ...)$ of being saturated. Here p is considered to be function of temperature T, concentration c etc.

Gels III





Figure 15: Crosslinking

Take the generating function

$$F_0 = \sum_n \omega_n^{(0)}(p)\theta^n \tag{89}$$

where $\omega_n(p)$ is the probability that a randomly picked monomer (A) is part of a cluster of n + 1 monomers. θ is a factor for every monomer in the cluster except for the considered monomer.

Now



$$F_0 = [1 - p + p\theta F_1(p, \theta)]^f = [(1 - p)u(p, \theta)]^f$$
(90)

where F_1 (with $\omega_m^{(1)}$ being the function corresponding to the situation that monomer B, attached to monomer A at the ν -th valence of A, is attached to *m* other monomers.

To make progress on the computation we assume that the f bonds of A are un-correlated and that the network has no loops (Cayley-tree, see Figure 16) (thus a mean-field approximation). Hence



Figure 16: The Cayley tree is a tree with all nodes having the same connectivity, here q = 3 and without loops

Gels V



$$F_{1}(p,\theta) = [1 - p + p\theta F_{1}(p,\theta)]^{f-1} = [(1 - p)u(p,\theta)]^{f-1} .$$
(91)

Solving this we find F_0 and find all $\omega_n^{(0)}$

$$\frac{1}{p\theta(1-p)^{f-2}}\left(\frac{p\theta}{1-p}F_1\right) = \left(1 + \frac{p\theta}{1-p}\right)^{f-1}$$
(92)

or

$$u - 1 = xu^{f - 1} . (93)$$

Thus the function $u = 1 + F_1 p\theta/(1-p)$ is a function of $x = p\theta(1-p)^{f-2}$ alone. One can calculate the Taylor-expansion of u

$$u = \sum_{n=0}^{\infty} \frac{[(f-1)n]! x^n}{[(f-2)n+1]! n!}$$
(94)

from which we find
Gels VI



$$F_1 = (1-p)^{f-1} \sum_{n=1}^{\infty} \frac{[(f-1)n]!}{[(f-2)n+1]!n!} x^{n-1}$$
(95)

$$F_0 = (1-p)^f f \sum_{n=1}^{\infty} \frac{[(f-1)n]!}{[(f-2)n+2]![n-1]!} x^{n-1} .$$
(96)

To understand F_1 we go back to Equation 91 and write

$$F_1 = w^{f-1} (97)$$

$$\frac{w-1}{p\theta} = w^{f-1} - 1/\theta .$$
 (98)

Since w is a monotonically increasing function of θ we find that only one intersection is possible. Assume $\theta = 1$, then there are two cases. Case one

$$f - 1 < 1/p$$
 or $p < \frac{1}{f - 1}$. (99)

Gels VII



This mean that at monomer B the average number of bonds is

$$p(f-1) < 1$$
. (100)

The chain forming dies out

$$\sum \omega_n(p) = 1 \tag{101}$$

and we only find finite connectivity. The system is in the sol phase. If

$$f - 1 > 1/p$$
 or $p(f - 1) > 1$ (102)

the from generation to generation the number of bonds increase. The *gel point* in this theory is thus

$$p_c = \frac{1}{f - 1} \,. \tag{103}$$

We shall now investigate the behaviour of the system in the neighbourhood of the gel point. To do so, we will expand various property function around

Gels VIII



$$\Delta = p - p_c = p - \frac{1}{f - 1} .$$
 (104)

The gel fraction is given by

$$G = 1 - F_0 = 2f \frac{f-1}{f-2} \Delta \sim \Delta^{\beta} , \quad \beta = 1 .$$
 (105)

Next we look at the polymerization index N_ω

$$N_{\omega} = \frac{\sum_{n} n \omega_{n}(p)}{\sum_{n} \omega_{n}(p)} = \frac{\partial}{\partial \theta} \ln F_{0}(\theta = 1) .$$
 (106)

We have

$$F'_0 = pf(1+F'_1)F_1$$
(107)

$$F'_{1} = p(f-1)(1+F'_{1})F_{1}^{(f-2)/(f-1)}$$
(108)

and for $\theta = 1$, $p < p_c$, i.e., $F_0 = F_1 = 1$

Gels IX



$$N_{\omega} = \frac{pf}{1 - p(f - 1)} \approx \frac{f}{|\Delta|} \sim |\Delta|^{-\gamma} .$$
(109)

The next observable we investigate is the cluster size distribution. With the help of the Stirling formula we find from Equation 96

$$\omega_n \sim e^{-nq(\Delta} n^{-3/2} \sim n^{3/2} e^{-n\Delta^{1/\sigma}}$$
(110)

with

$$q(\Delta) = -(f-2)\ln\left(1 - \frac{f-1}{f-2}\Delta\right) - \ln[1 + (f-1)\Delta]$$
(111)
$$1(f-1)^{3} + 2$$

$$\approx \quad \frac{1}{2} \frac{(r-1)}{f-2} \Delta^2 \,. \tag{112}$$

All in all we find the typical signature for a second-order phase transition with critical exponents β,γ,σ,τ given by

Gels X



$$G \sim \Delta^{\beta}$$
 , $\beta = 1$ (113)

$$N \sim |\Delta|^{-\gamma}$$
, $\gamma = 1$ (114)

$$\omega_n \sim n^{-\tau} e^{-n|\Delta|^{1/\sigma}}$$
, $\tau = 3/2, \sigma = 1/2$. (115)



Excercises

Excercises I



Exercise 1: Erdös-Renyi-Model with fitness

Code the following algorithm

Algorithm 2 Barabasi-Albert-Model

- 1: Start with a disconnected set of *m* vertices, each characterized by a constant ability (fitness) η_i to attract new edges. Assume a distribution $\rho(\eta)$
- 2: As in the Barababsi-Albert-model there is growth since vertices enter the system
- 3: There is preferential attachment

$$\Pi(k_i,\eta_i) = \frac{k_i\eta_i}{\sum_{j=1}^n k_j\eta_j}$$

Exercise 2: Watts-Strogatz Model

The Watts-Strogatz graph model is generated as follows:

- Construct a ring lattice with *N* nodes of mean degree 2*K*. Each node is connected to its *K* nearest neighbors on either side.
- For each edge in the graph, rewire the target node with probability *p*. The rewired edge cannot be a duplicate or self-loop.

Excercises II



What happens at p = 1?

Exercise 3: Network properties

Compute the local and global clustering coefficient of the following networks:

- finite square lattice
- finite heaxagonal lattice
- Infinite two-dimensional lattice, where each node is connected to its 8 closest neighbors
- Exercise 4: Develop an algorithmic test that, given an undirected network represented by its adjacency matrix, determines whether or not the input network is likely to be a real-world network.



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